

DIGITALIS **PROJECT**

"anything of lasting value will, once digitized and spread on the world wide web, exist and be available to all until the end of the Technological Era."

THE DIGITALIS PROJECT:

the Psilocybin Producer's Guide

by adam gottlieb

Copyright © 1976 by Kistone Press

Cover and illustrations by Larry Todd

TABLE OF CONTENTS

Introduction		 	 	 	 					:	2
Psilocybin and the Law		 	 	 	 					:	2
Finding the Mushroom		 	 	 	 					:	3
Data on Various Psilocybian Species		 	 	 	 					'	4
Pure Culture Technique		 	 		 	 				'	7
Sterilization		 	 	 	 						8
Making a Spore Print		 	 	 		 				:	8
Preparation of Media											
Equipment		 	 			 				1	0
Starting the Culture		 	 			 				1	0
Raising Crop Cultures of Mycelia		 	 			 				1	11
Harvesting and Drying		 	 							1	2
Extraction		 	 							1	12
Dosage		 	 							1	3
Large Scale Production		 	 							1	3
Maintaining a Perpetual Psilocybin Fari	m	 	 							1	6
Recommended Reading		 	 				 			1	6

INTRODUCTION

If a person knows what he is doing, it is not difficult to cultivate the mycelium of any of the psychoactive psilocybin bearing mushrooms. The mycelium is the fibrous underground network of the mushroom. The familiar stem and cap portions of the mushrooms are called carpophores. The mycelium can be readily grown in ordinary Mason jars in a low cost medium in 10 to 12 days and the active materials (psilocybin and psilocin) can be easily extracted. This book explains how to carry out all of these steps on a small or large scale. Co.nplete instructions are given for locating the mushrooms, developing stock cultures for inoculation, cultivating, harvesting, and drying the mycelium, extracting the active alkaloids, and using the existing cultures to seed new cultures to keep an ongoing psilocybin farm yielding a regular crop of the hallucinogenic mycelium. We also give directions for setting up in a small workroom a large scale psilocybin factory which can product at least 5,000 doses of the drug every week.

PSILOCYBIN AND THE LAW

The present drug laws are a pathetic mess. The old adage that igorance of the law is no excuse becomes a ludicrous statement when the laws themselves are rooted in ignorance. One classical example of this is the classification of the stimulant cocaine as a narcotic. One is reminded of the king in Alice in Wonderland who made up his own language as he went along with total disregard for the accepted definitions of words. I will not even go into the question of whether any law enforcement agency has the moral or Constitutional right to dictate what substances we may or may not take into our own adult bodies. Any modern individual whose mind is not immersed in the slavish dung pit of Dark Age unreasoning knows that reliable education — not criminal penalization — is the answer to whatever drug problems exist. Nevertheless, we must contend realistically with the powers that unfortunately be at this time. They are the ones with the badges, guns, gavels and goons.

Because of the afore mentioned ignorance of our lawmakers it is difficult to determine how the use of certain hallucinogenic substances would be treated in the courts. Possession of psilocybin and psilocin (misspelled in the U.S. Code as psilocyn) is a felony under Title 21, Section I, (C) of the United States Code (1970 Edition). Psilocybe mexicana is also illegal. There was sufficient ignorance on the part of the law makers not to include the many other mushroom species containing psilocybin

and psilocin. Theoretically the possession of any psilocybin-bearing mushroom would be the same as possessing the alkaloid itself. But when it comes to prosecution it does not necessarily work like that. Lysergic acid amides, which occur in morning glory seeds, stems, and leaves are also illegal, but there is no way to prevent gardeners from raising this ornamental flower. It is illegal for anyone in the USA to possess mescaline. Pevote, which contains mescaline, is legal for bonafide members of the Native American Church when used ritualistically, but no member may possess extracts of the cactus or the drug mescaline. Peyote is illegal for non-members, but San Pedro and several other species of Trichocereus cacti also contain mescaline and are available from many legitimate cactus dealers. It would be clearly illegal for anyone to extract the active principles from any of the above mentioned plants. And it would be illegal for anyone to extract psilocybin and psilocin from mushrooms or mycelium as described in this book. Anyone found operating a large scale mycelium farm could very easily be prosecuted for intent to manufacture psilocybin and psilocin. There are also many different state laws which must be considered before doing anything with psilocybin-bearing mushrooms. There are, however many nations which have no laws regarding these substances. We are not judges or attorneys and are not trying to offer clear interpretations of the law. Rather we have mentioned these points to give some indication of the legal pitfalls which surround the application of the activities described in this book. Furthermore, laws are constantly being revised. By the time this book is published and read the laws may have changed for better or for worse. We, the author and publisher, are not recommending or endorsing the application of the information in this book especially in places where there are laws proscribing these substances. We offer this information for the sake of pure knowledge because it is our constitutional right to do so. We do not encourage the violation of any existing laws.

FINDING THE MUSHROOM

All it takes is one mushroom or a few spores and from this one can quickly develop a culture that will continue to produce as much psilocybin as one desires for years to come. Because the common San Ysidro mushroom *Psilocybe cubensis* Singer (formerly *Stropharia cubensis* Earl) is the most easily obtainable, most readily cultivated, most disease resistant, and psychoactively strongest species we have geared our instructions to its use. There are, however, numerous other species which contain psilocybin. In case one of these is all that is available, we give for several of these pertinent information such as relative potency, where and when to find specimens, what growing conditions (medium, temperature, lighting, etc.) it favors and how resistant it is to contamination. The states, provinces, and regions named are by no means the only places where the species is to be found. They are places in which there have been numerous reports of findings. They are given here to give a general idea of the type of terrain and climate the species favors. In cases where ideal cultivation temperatures and growing conditions are not given much can be surmised by considering the environment in which that species thrives.

Psilocybe cubensis can be found in many parts of the United States, Mexico, Colombia, Australia, and even Southeastern Asia. It is usually found growing on or near cow dung in pastures during warm rainy periods from February to November.

There are several species of mushroom which occur on cow dung, but fortunately none of these bears much resemblance to the San Ysidro.

There are numerous toxic mushrooms growing around us. Some of these could be mistaken for some of the other psilocybin fungi mentioned in this book. It is essertial that the mushroom hunter learn to use an identification key. A key is a listing of various features which will positively identify a given species. If a specimen does not conform in every respect to the key, it must not be used. There are several excellent keys to be found on most library shelves. One that we recommend is *Keys to Genera of Higher Fungi* by R. Shaffer. 2nd ed. (1968) published by the University of Michigan Biological Station at Ann Arbor. We also recommend a thorough reading of a most helpful book, *Poisonous and Hallucinogenic Mushrooms* by Richard and Karen Haard, available for \$3.95 from Nature Study Institute, P.O. Box 2321, Bellingham, Washington 98225. It is further suggested that after identifying the specimen it should be brought to an expert mycologist to be absolutely certain of its identify.

Many books on hallucinogenic mushrooms suggest a simple test for psilocybian species which involves breaking the flesh of the specimen and waiting about 30 minutes for a bluing reaction to take place. This bluing is due to the oxidization of indole based substances in the fungus. Although it is true that most of the psilocybin-bearing mushrooms will respond positively to this test, other species may do the same. The poisonous Eastwood Boletus blues upon exposure of the inner tissues to oxygen as well as does any psilocybin mushroom. Antoher test which is often given in mushroom manuals in treating the exposed tissues with Metol a chemical used in photo developers. It hastens the bluing of psilocybian mushrooms, and supposedly one can do a bluing test with it in a few minutes that would otherwise take 30 minutes or more. Any mushroom, however, which contains indolic substances of any sort will respond positively to this test. Since indole-based amino acids such as tryptophan are found in most living organisms this test is rather useless.

There is actually no field test for psilocybian mushrooms. There is however, a relatively simple test for the presence of psilocin and psilocybin that can be carried out at home by anyone who has some familiarity with paper chromatography. The mushroom sample is dried, pulverized, and extracted into a small amount of unheated methanol by shaking for half an hour. After the debris in the methanol has settled the paper is spotted with the top fluid in a zone about 2mm. After treating the spotting zone with water saturated butanol for about 2 hours, the solvent front 7–8 cm from the spotting zone would contain the psilocin and psilocybin if they were present in the specimen. After drying the paper with a hair dryer on warm this outer zone is sprayed lightly with a saturated solution of p-dimethyl-aminobenzaldehyde in alcohol and then again with 1 N hydrochloric acid. The paper is then dried again as before. Where psilocybin is present a reddish color will develop. The presence of psilocin will be indicated by a blue-violet zone.

DATA ON VARIOUS PSILOCYBIAN SPECIES

Conocybe cyanopes: Found from May through September usually in dense shade scattered among mosses, and in wet soil around bogs, swamps, and ditches in north-

western USA and as far east as Michigan. Carpophores grow well in sphagnum moss having a range of pH 7-8.

Copelandia cyanescens: Found in early summer through late autumn scattered, grouped, or clustered on cow dung, or rich soil in Florida and other southern states. Spores germinate easily on all agar media. Optimum growth occurs on MEA at 80 degrees F. Carpophores can be produced on uncased compost or on rye.

Panaeolus foenisecii (also known as Paneaolina foenisecii or Psilocybe foenisecii, and commonly known as haymower's mushroom or harvest mushroom): found in late spring and early summer, or in July, August, and September during cool, wet seasons scattered or grouped in large numbers on lawns, pastures and other grassy places throughout the USA and in Quebec. Tests on specimens found in Washington revealed no psilocybin, but eastern specimens were potent.

Panaeolus sphinctrinus: Found in summer and autumn in small groups in forests, pastures, fields, and roadsides almost always on cow dung in many temperate parts of the world.

Panaeolus subalteatus: Found from spring to autumn grouped or clustered often in rings up to two feet in diameter on open ground, freshly manured lawns, straw piles, all types of compost, dung piles, and roadsides in Ontario and throughout the USA (especially in Massachusetts, Maryland, New York, Ohio, Michigan, Washington, and Oregon). Optimum growth in MEA is at 86 degrees F. It occasionally occurs as a weed mushroom in commercial mushroom houses.

Pholiotina cyanopoda: Found from August through September solitary to clustered on lawns in such diverse parts of the USA as New York, Washington, and Colorado.

Psilocybe baeocystis: Found in autumn and winter, solitary, grouped or clustered on earth, lawns, mulch, and decomposing forest wood near scattered trees especially conifers — in western Oregon and Washington. It does well on all agar media at 77 degrees F. This is a potent species containing psilocybin, psilocin, baeocystin and nor-baeocystin. Perhaps it is because of the latter two alkaloids that it is the most visually hallucinogenic of the psilocybian mushrooms. There is a report that in 1960 a six-year old boy died after eating a large number of these mushrooms. There has never been any other indication that these alkaloids are dangerous. Until there is further clarification of this question we advise that anyone using this species proceed with caution by starting with small doses and progressing gradually to larger ones. This is especially important when using the extracted crude alkaloids which may contain large concentrations of the baeocystin alkaloids.

Psilocybe caerulescens: Found in summer during rainy season, grouped or clustered, but rarely solitary, mostly in shady places on soil, sugar cane mulch, recently turned earth or stream banks — in Alabama, northern Florida and Mexico. The Mexican variety P.caerulescens var. mazatecorum is known locally as derrumbe, which means "landslides." There it is often found among landslides, or near corn or coffee plantations. The mycelium does best on MEA at 81 degrees F. Thermal death occurs at 95 degrees F. It is almost impossible to produce carpophores on sterilized rye medium. They can be grown on vegetable compost in dim light, but the incubation period is long (55–85 days). Although this species is resistant to white mold its long incubation period leaves it prone to other diseases. It is not one of the more potent species.

Psilocybe caerulipes: Found in summer and occasionally autumn solitary or clustered on decomposing logs and debris of hardwood trees (especially birch and maple) in New York, New England states, Ohio, Michigan, North Carolina, Tennessee and Ontario.

Psilocybe cubensis var. cyanescens Singer (formerly Stropharia cubensis Earl): Found from February to November in compact groups in clearings outside forest areas, on cow dung, or horse dung, in rich pasture soil, on straw, or on sawdust/dung

mixture in Mexico, Cuba, Florida and other southern states. It grows well on MEA. At 86 degrees F carpophores appear in 4–8 weeks. Thermal death occurs at 104 degrees F. Carpophores larger than wild specimens can be produced by inoculating vegetable compost in clay pots with agar grown mycelium, casing with silica sand/limestone mix, and incubating 4–6 weeks in daylight at 68 degrees F. It does poorly in darkness. It is a potent mushroom and is very resistant to contaminants.

Psilocybe cyanescens: Found in autumn scattered, grouped or clustered in woods, on earth, among leaves and twigs, and occasionally on decomposing wood — in northwestern USA.

Psilocybe mexicana: Found from May to October isolated or sparsely at altitudes from 4500 to 5500 feet, especially in limestone regions, among mosses and herbs, along roadsides, in humid meadows, in cornfields, and near pine forests in Mexico.

Psilocybe pelliculosa: Found September to December scattered, grouped, or clustered on humus and debris, in or near conifer forests in north-western USA and as far south as Marin County, California. This is a small but potent species.

Psilocybe quebecensis: Found from summer to late October scattered in shady areas at forest adges, on sandy soil containing vegetable debris regularly inundated by river flooding, and on decomposing wood and debris (especially birch, alder, fir and spruce) in the Quebec area. It thrives at lower temperatures than other Psilocybe species and produces carpophores at air temperatures of 43 to 59 degrees F.

Psilocybe semilonceata: -ound August through Setpember often in large groups on soil, among grasses, in clearings, pastures, meadows, forest edges, open conifer woodlands and on roadsides — but never on dung — in New York, northern USA, British Columbia, and Europe. Generally regarded as one of the less potent species, but is sometimes quite potent.

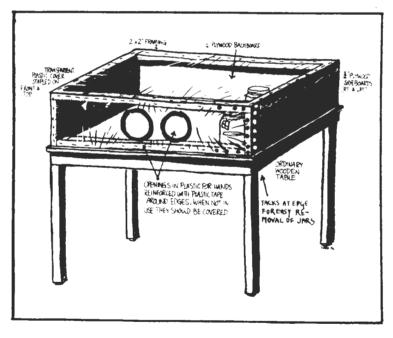
Psilocybe strictipes: Found in October rather clustered on soil, or on decomposing wood and debris or conifers and some other trees in north-western USA (especially in Oregon). It closely resembles *P.baeocystis*, but has a longer stem. It tends to be as visually hallucinogenic as that species and probably contains the same or similar baeocystin alkaloids.

Psilocybe sylvatica: Found in September and October in small compact but unclustered groups in woods on leaf mold, on debris (especially beech wood), around stumps and logs, but not usually on them — from New York to Michigan and as far north as Quebec and Ontario. This mushroom is small and is often mistaken for P. pelliculosa.

The species discussed above are only some of the more commonly known ones with hallucinogenic properties. There are recognized among the psilocybin-bearing mush-rooms 40 species of Conocybe usually occuring in forests, pastures, gardens, dung areas, sandy soil, ant hills, decayed wood, and charcoal and having a cosmopolitan range; 20 species of Panaeolus found on soil and dung and having a cosmopolitan range; 40 species of Psilocybe found on soil, moss clumps and organic substrata such as dung, rotting wood, bagasse, and peat ranging from the arctic to the tropics; and 9 species of Stropharia found on soil, dung and sometimes on leaf mulch and rotting wood and having a fairly cosmopolitan range.

PURE CULTURE TECHNIQUE

The most difficult part of psilocybian mushroom cultivation is the observance of the rules of pure culture technique. These are the sanitary code of mushroom cultivation. There are usually many varieties of bacteria and fungal spores in our environment; floating in the air, clinging to our hands and clothing, issuing from our mouths with every exhalation. Extreme measures must therefore be taken to keep these out of our mycelial cultures, which they could rapidly overrun. The following points should be diligently observed. Work in a clean, uncluttered, dust-free room. Immediately before work wash the work table and spray the room with disinfectants. Scrub arms, hands and nails with disinfectant soap. Wear simple clothing. A freshly cleaned shortsleeve T-shirt is ideal. Gargle with antiseptic mouthwash and cover the mouth and nose



Home made inoculation hood.

with a clean cloth or disposable surgery mask. Cover the hair with a surgical cap or shower cap. Allow no drafts in the room. Close all windows and stuff all door jambs. Let no flies, animals or unnecessary people in the room. Let only sterilized equipment touch the medium or inoculum. Don't lean over your work. Avoid all swift in exempts that may cause a draft. If possible have a hood constructed around the work table or a screen or curtain surrounding it. Be neat and keep all materials within reach. Keep all equipment about three feet away from the work. Do not permit anyone to enter or exit the room while work is in progress.

STERILIZATION

All otensils used in the cultivation of the mycelie must be sterilized by heat before use. Glassware must be boiled in water for 30 minutes. Metalware used repeatedly must be held in a flame until glowing and then allowed a moment to cool before making contact with any cultures or specimens. When the inoculation loop has been used to transfer a fragment of mycelium it must be flame sterilized again before touching the next fragment. All medium containers must be sterilized after the medium has been poured. This process is known as autoclaving. Containers no more than half full or th medium are placed in a canning type pressure cooker. The lids of these must be loose enough to allow escapage of internal pressures. Otherwise the containers may crack. Seal the lid of the pressure cooker. Keep the stopcock walve open. Using high heat, bring the cooker to boiling so that thick steam comes through the vent. Close the stopcock and let the pressure rise to 15-20 lbs. (250 degrees F) for 30 minutes. This should be enough to destroy any foreign spores or life-forms. Any higher temperature or longer period could cause the dextrose or maltose sugara to carmelize. This would inhibit growth and psilocybin production of the mysulium. When the autoclave period is up turn off the heat and let the cooker cool to room temperature. Do not release the stopcock until everything has thoroughly cooled or the sudden change in pressure will cause the containers to boil over. Discard any containers that have cracked during sterilization. Keep all containers of medium at room temperature for three days to see if any foreign molds develop. If they do occur discand the medium in the contaminated jars and thoroughly clean and sterilize such jars before using again.

MAKING A SPORE PRINT

A spore print is a collection of spores on a flat surface. It can serve several purposes. It can be used to essist identification of the specimen by observing its color or if made on a glass slide, by studying the shapes of the spores under a microscope. Mycological identification keys include descriptions of spore prints and microscopic spore features for different species. Spore prints are also the standard method of collecting spores for

later germination on again media. A print from a single mushroom cap contains millions of spores. Many mushroom lovers are now making spore prints on paper from species available in their locales and mailing them to cultivators in other areas where such species are not found. Secret spore exchange correspondence clubs are becoming quite the vogue and will probably be more common in the very near future. A word of caution regarding this practice should be given, however. Do not assume that spores received in this manner are from the species that the sender claims they are. If the sender has misidentified the specimen and the recipient cultivates and ingests mycelia or extractions therefrom, the result may be disasterous. Furthermore, I would not put it past some anti-drug fanatic to purposefully disseminate spore prints of dangerous mushrooms to amateur culvitators. This could result in sickness and death for thousands of persons.

To make a spore print take a mushroom with its cap fully opened and gills exposed. With a sharp sterilized blade cut off the stem as close to the gills as possible. Place the cap gills down on a clean, white sheet of paper, or on a sheet of glass that just been swabbed with alcohol, or on two or four sterilized microscopic slide glasses. Cover the cap with a clean, inverted bowl or bell jar to prevent drying of the cap and intrusion of foreign organisms. Let this stand as such for 24 hours. If a good spore print has not formed after this time, tap the cap lightly with the flat side of a knife or spatula. This should shake loose many spores. If the print is made on glass, cover it with another glass sheet immediately after removing the cap to prevent contamination. If microscopic slides are used, place two face to face and seal the edges with tape. If paper is used, fold it several times so ther she print is well inside.

PRÉPARATION OF MEDIA

PDA (Potato Dextrose Yeast Ager): Wash 250 grams of unpeeled potatoes and slice thum 1/8 inch thick, Wash these several times in cool tap water until the water is clear. Drain the slices in a collander and rinse once with distilled water. Cook the potato slices in distilled water until tender. Strain the cooking liquids through a flannel cloth or several layers of cheesecloth and collect the liquid in a flask. Rinse the potatoes several times with distilled water, add these rinse waters to the liquid in the flask, and discard the potatoes. Add enough distilled water to the flask to make one liter. Bring the liquid to a boil and add 15 grams of agar, 10 grams of dextrose, and 1.5 grams of yeast extract. The agar must be added slowly and carefully to prevent boiling over. While the liquid is hot pour it into petri dishes or other culture containers. Each should be filled about half-way.

PDY broth is made in exactly the same manner except the agar is omitted. Mason jars are filled half way with the hot or cool liquid.

MEA (Malt Extract Agar): To one liter of gently boiling water (distilled) add 20 grams of malt extract, 20 grams of agar (slowly, carefully to prevent boiling over), 100 mg of potassium phosphate dibasic (K₂HPO₄), and 100 mg of calcium carbonate. While still hot pour the liquid into the culture dishes.



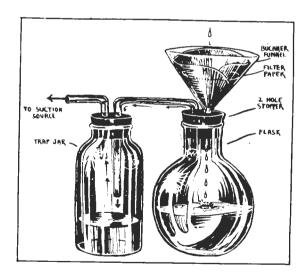
4

HARVESTING AND DRYING

Filter the medium of each jar through a clean flannel cloth, collect the mycelial material from the cloth, and place it in a pyrex baking dish. Do the same with each jar of mycelium until each baking dish is about 1/3 full with mycelia. Dry these in an oven at no higher temperature than 200 degrees F. Use an oven thermometer. Do not rely upon the temperature indications on the oven knob as these may vary from accuracy. Check the baking dishes periodically. When the material first appears to have died shut off the neat and let the dishes stay in the oven until it has cooled. This ensures the evaporation of residual moisture. Each cultivation jar should yield 50–100 grams of wet mycelium. Fresh mycelium contains about 90% water, so this much would dry down to 5 to 10 grams of crumbly material. Each baking dish would contain a dozen or more mycelia.

EXTRACTION

Crumble and pulverize the dried mycelial material and combine each 100 mg of this with 10 ml of absolute methanol. Place the flask in a hot water bath for four hours. Filter the liquids with suction through a filter paper in a buchner funnel with Celite



Suction filter.

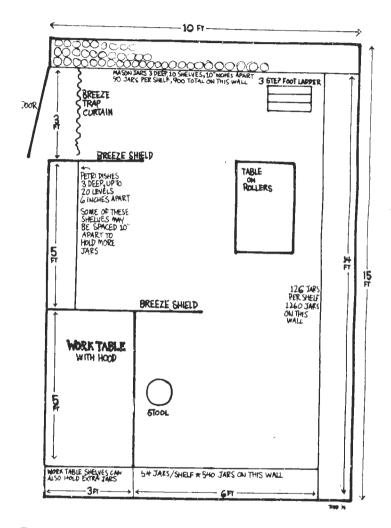
to prevent clogging. Collect and save the filtrate liquids. Heat the slurry (the mush in the filter paper) two more times in methanol as before, filter, and accumulate the liquids of the three extractions. To be certain that all of the alkaloids have been extracted do a small extraction of a portion of the used slurry and test with Keller's reagent (glacial acetic acid, ferrous chloride, and concentrated sulfuric acid). If there is a violet indication, alkaloids are still present and further extraction is in order. In an open beaker evaporate the liquids to total dryness with a hot water bath or by applying a hair dryer. Be certain that all traces of methanol have been removed. The remaining residue should contain 25-50% psilocybin/psilocin mixture. Greater purification can be achieved, but would require other solvents and chromatography equipment and is hardly necessary. Each 100 grams of dried mycelium should yield about 2 grams of extracted material. This should contain at least 500 mg of psilocybin/ psilocin mixed or about fifty 10 mg doses. Theoretically psilocin should have the same effect upon the user as psilocybin. The only difference between the two is that the latter has a phosphate bond which disappears immediately after assimilation in the body. In other words, in the body psilocybin turns into psilocin. Psilocybin is a fairly stable compound, but psilocin is very susceptible to oxidization. It is best to keep the extracted material in a dry air tight container under refrigeration. A sack of silica-gel can be placed in the container to capture any moisture that may enter.

DOSAGE

The standard dose of psilocybin or psilocin for a 150 lb. person is a 6–20 mg. We will figure the average dose as 10 mg. The crude alkaloid extraction process given here yields a brownish crystalline powder that is at least 25% pure. Each Mason jar should contain at least 50 grams of wet mycelium. After drying this would be about 5 grams of material. The crude material extracted from this should contain 25–35 mg. of psilocybin/psilocin or roughly 2–3 hits. This yield may vary to some extent depending upon several factors. Many species contain less of these alkaloids than does psilocybe cubensis and the alkaloidal content of this species may vary in different strains. Cultivation conditions have a lot to do with yield too. Higher temperatures (75 degrees F) cause more rapid growth but lesser psilocybin content than do lower temperatures (70 degrees F). One must test each new batch of extracted material to determine the proper distribution of dosages. Depending upon the potency of the mycelia and how well the extraction was conducted the dose may range between 25 and 100 mg. Also bear in mind that the dose varies for different individuals.

LARGE SCALE PRODUCTION

The techniques and procedures described in this book can be employed to cultivate modest supplies of psilocybin for personal use, or they can be expanded to apply to the large scale production of many thousands of doses per week. The diagram on the following page shows one way in which a small 10 x 15 foot room with standard 8 foot ceilings can be set up to produce an unending yield of at least 5000 doses per week.



The stock culture shelves here are 1 foot deep and 5 feet long. Each could hold twenty 15 cm petri dishes. If the shelving is spaced six inches apart, there can be as many as 16 shelves stacked in this space. This would allow for up to 300 stock culture dishes going at one time. The crop culture shelves can be stacked ten inches apart, accommodating one quart size Mason jars and giving ten levels. With the dimensions of the room depicted in this diagram this much shelving could hold about 2800 jars (3 deep and 3 per running foot). The entire room — walls, ceiling and shelving — should be painted with a white, glossy kitchen enamel. This is not only an important sanitary measure, but also improves the efficiency and even distribution of light in the

room. Lighting should be provided by a few banks of wide spectrum fluorescent tubes fairly evenly distributed across the ceiling and turned on for 10-12 hours regularly each day. These are great dust catchers, however, and must be wiped clean periodically. The work table should also be painted with a hard, smooth, white finish. If the table is of metal, a small, clean cutting board must be provided on which to pin down mushroom caps when disecting them. Shelf boards on the wall left of the table may extend above the table to provide space for storage of work equipment and readv containers. A hood should be constructed around the table to protect this space from dust, etc. A fume hood with a flu vent and spark-free exhaust fan should be constructed over the extraction table to remove toxic and combustible methanol vapors. Extraction is preferably conducted in another room. If the cultivation room is used for extraction while cultures are growing, care must be taken that the heat from the extraction processes does not alter the room temperature. The fume hood will help by carrying off much of the heat. A vinyl shower curtain should be hung to the right of the table to shield the work area from breezes when anyone enters or exits the room. Another vinyl curtain can be hung just inside the entrance to serve as a dust trap. A person entering would close the door behind him before pulling the curtain aside and visa versa on exiting. The floor should be white vinyl or asphalt tile or painted white and coated with verathane or polyurethane. There should be no cloth or carpeting in the room except for a supply of clean work clothing and surgical masks. The only other items in the room would be a stool at the work table, a three-step ladder for reaching the upper shelves, and a small table on rollers on which to place jars and dishes when making the rounds of the shelves.

Unless one has a large staff of assistants it would be impossible to inoculate 2800 jars in one work session. After getting used to the work one could do about 100 jars an hour. The best procedure is to set up a continuous rotation of inoculations. Working about 3 hours a day about 235 jars could be inoculated each session. All 2800 jars could be inoculated in 12 days. Sections of shelving would be divided into groups of 235 jars, and these sections would be labled with the date and approximate time of inoculation. The work schedule for cultivation would be as follows:

Day Inoculate		Shake	Harvest	Reinoculate				
Mon.	Group A							
Tues.	В							
Wed.	С	Group A						
Thurs.	D	В						
Fri.	E	A & C						
Sat.	F	B & D						
Sun.	G	A, C & E						
Mon.	·H	B, D & F						
Tues.		A, C, E & G						
Wed.	J	B, D, F & H						
Thurs	K	A, C, E, G & I						
Fri.	L	B, D, F, H & J						
Sat.	Commence Reinoculation see Col. 5	C, E, G, I & K	Group A	Group A-2				
Sun.		D, F, H, J & L	В	B-2				
Mon.		E, G, I, K & A-2	С	C-2				
Tues.		F, H, J, L & B-2	D	D-2				
etc.		etc.	etc.	etc.				

This would represent the first two weeks of the continuous cultivation cycle. The continuation of this schedule is obvious: shaking every other day, harvesting approximately every 12 days, and resterilizing, refilling with fresh medium, autoclaving, and reinoculating the jars liberated by the day's harvest. If the total number of jars is 2800, each group would consist of about 235 jars. This same schedule could, of course, be adapted to any total number of jars. Drying of mycelia should be done within a few hours after harvesting. Otherwise enzymes in the material will begin to destroy the active alkaloids. Once dried the material can be stored in a cool, dark, dry place until enough daily harvests have been accumulated to do an extraction. If the mycelia can not be dried right away it can be kept in a refrigerator for a day or two, or for longer times in a freezer.

MAINTAINING PERPETUAL PSILOCYBIN FARM

Fresh inoculum can come from stock culture dishes kept under refrigeration. If these should become depleted, healthy strains of mycelium from the crop cultures can be used to inoculate sterilized agar media in the dishes. To do so shake the crop culture jar violently to break up the mycelium. Then transfer drops of the liquid to autoclaved petri dishes of unused agar medium with a sterilized pipette and let it grow as before. If this reinoculation of stock cultures from existing crops is continued over a long period of time, the strain will eventually weaken due to what is known as the senescence factor. To avoid this alternate the media used in the stock dishes. That is: if PDA is used the first time, use MEA the second time and PDA again the next time, etc.

RECOMMENDED READING

If you can not find these books in a bookstore, they can be ordered by mail from their publishers.

Home Grown Highs. M. J. Superweed, 1972. High potency cultivation techniques for several psychoactive plants including peyote, San Pedro, coleus, and morning glory; plus a special medium formula and practical method for maximum mycelial growth and extra high psilocybin yield. The formula can be used in combination with the large-scale production methods in our book. Send \$1.50 plus .50 handling to: Flash Mail Order Post Express, 9926 Haldeman Ave., Suite 3B, Philadelphia, Pa. 19115.

Psilocybin—Magic Mushroom Grower's Guide. O. T. Oss and O. N. Oeric, 1976. Excellent guide for those who wish to cultivate carpophores of Stropharia cubensis. Nicely illustrated with black and white, and color photos. Send \$4.95 plus \$.50 handling to: Flash Mail Order Post Express, 9926 Haldeman Ave., Suite 3B, Philadelphia, Pa. 19115.

Two other highly recommended books — Keys to Genera of Higher Fungi and Poisonous and Hallucinogenic Mushrooms are discussed on page 4 of our book.

DIGITALIS **PROJECT**

"anything of lasting value will, once digitized and spread on the world wide web, exist and be available to all until the end of the Technological Era."

THE DIGITALIS PROJECT